

The effect of environmental pollution on the American cockroach in Iraq for the genes COX1, COI LCO

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ABSTRACT

This research aims to determine the effect of environmental pollution on some genetic markers of the American cockroach by conducting molecular tests on the COX1 and COI LCO genes. The current study was conducted at Samarra University, College of Education/Laboratory of the Department of Life Sciences. Samples of adult American cockroaches were collected from six Iraqi cities: Duhok, Kirkuk, Samarra, Baghdad, Nasiriyah, and Basra, between September 2023 and September 2024. Three samples were taken from each city for molecular study. DNA was extracted, polymerase chain reaction (PCR) was performed, and the samples were sent for DNA sequencing analysis. The results showed 100% similarity between samples in the DNA analysis of the COX1 gene, indicating that this gene is suitable for studying convergence between species. However, the COI LCO gene showed interspecies differences and is suitable for studying environmental effects.

1. INTRODUCTION

Cockroaches are one of the most widespread insects in the world, comprising more than 5,000 species. They are considered urban pests that cause harm to humans, such as disease transmission and food spoilage. Cockroaches can live in all locations and environments. They can live in both hot and cold regions, but prefer warm environments. The American cockroach, one of the most widespread insects in the order of cockroaches, is known for its low mobility and is most active at night. In the dark, they cease movement and tend to remain dormant (Nguyen et al., 2020; Ademolu et al., 2020). The American cockroach is one of the most widespread insects in Iraq. Originally from South America, it is 40 mm long, reddish-brown in color, and lives in warm areas such as kitchens, heating rooms, warehouses, and sewage systems. It emerges at night for feeding and other activities. Long-lived cockroaches live for more than a year and are among the most annoying household pests, causing damage to household materials and stored products. They are characterized by their mixed diet, feeding on plants, meat, and food scraps. The American cockroach has a very high capacity for reproduction and adapting to different environmental conditions (Zhao et al., 2022; Cerreta et al., 2022). The environment is constantly changing, and survival under these conditions is difficult, especially for organisms such as insects, which play a major role in the long-term dynamics of ecosystems. Therefore, their extinction or decline in numbers will impact the human food chain and the processes taking place in agricultural soil, affecting crop quality. There are also insect species, such as bees, whose life cycles are affected by temperature changes. Furthermore, human practices, such as the use of Excessive pesticides and chemicals affect insects, in addition to the effects of global warming (Barton et al., 2017; Dominic et al., 2023).

Genes provide the most important information about insect development, including mitochondrial DNA (mtDNA), which in the American cockroach consists of a double strand ranging in size from 14,903 to 19,513 base pairs. This gene has been classified as an important taxonomic tool. Among these genes is the COI gene, considered one of the most important genes used in insect classification. It has several types and is a common region across all organisms, but differs in the distribution of nitrogenous bases. It is widely used in insect classification to determine similarities between species (Shi et al., 2021; Ma et al., 2018).

The American cockroach feeds on many species and is widespread in all environments. Therefore, it contains proteins, heavy and light nutrients, and even heavy elements, including cobalt, zinc, and lead (Boate et al., 2021; Hamanaka et al., 2016). This study aims to know the effect of environmental pollution on the American cockroach by studying the genes COX1 and COI LCO.

DNA extraction

Measure 25 mg of ground tissue sample, and then transfer into 1.5 ml tube using a spatula, Add 200 µl Buffer CL, 20 µl Proteinase K and 5 µl RNase A Solution into sample tube and mix by vortexing vigorously, Incubate the lysate at 56°C (preheated heat block or water bath) for 10 ~ 30 min, After lysis completely, add 200 µl of Buffer BL into upper sample tube and mix thoroughly. Then incubate the mixture at 70°C for 5 min, Centrifuge the sample tube at 13,000 rpm for 5 min to remove un-lysed tissue particles. Then carefully transfer 350 ~ 400 µl of the supernatant into a new 1.5 ml tube (not provided), Add 200 µl of absolute ethanol into the lysate, and mix well by gently inverting 5 - 6 times or by pipetting. DO NOT vortex. After mixing, briefly centrifuge the 1.5 ml tube to remove drops from inside of the lid, Carefully apply the mixture from step 6 to the Spin Column (in a 2 ml Collection Tube) without wetting the rim, close the cap, and centrifuge at 13,000 rpm for 1 min. Discard the filtrate and place the Spin Column in a 2 ml Collection Tube (reuse), Add 700 µl of Buffer WA to the Spin Column without wetting the rim, and centrifuge for 1 min at 13,000 rpm. Discard the flow-through and reuse the Collection Tube, Add 700 µl of Buffer WB to the Spin Column without wetting the rim, and centrifuge for 1 min at 13,000 rpm. Discard the flow-through and place the Column into a 2.0 ml Collection Tube (reuse), Then again centrifuge for additionally 1 min to dry the membrane. Discard the flow-through and Collection Tube altogether, Place the Spin Column into a new 1.5 ml tube (not supplied), and 30 - 100 µl of Buffer CE directly onto the membrane. Incubate for 1 min at room temperature and then centrifuge for 1 min at 13,000 rpm to elute.

Maxime PCR PreMix kit (i-Taq) 20µlrxn (Cat. No. 25025)

iNtRON's *Maxime* PCR PreMix Kit has not only various kinds of PreMix Kit according to experience purpose, but also a 2X Master mix solution. *Maxime* PCR Pre Mix Kit (*i*-Taq) is the product what is mixed every component: *i*-Taq DNA Polymerase, dNTP mixture, reaction buffer, and so on-in one tube for 1 rxn PCR. This is the product that can get the best result with the most convenience system. The first reason is that it has every components for PCR, so we can do PCR just add a template DNA, primer set, and D.W.. The second reason is that it has Gel loading buffer to do electrophoresis, so we can do gel loading without any treatment. It is suitable for various sample's experience by fast and simple using method.

Results and Discussion

Table (1) COX1 gene primers

Primer	Sequence	Tm (°C)	GC (%)	Product size
Forward	5'-GGAGCCCCAGATATAGCTTTCC - 3'	57	54.5	420 base pair
Reverse	5'-AAATTGGATCTCCCCCTCCTG - 3'	56.6	52.4	

Table (2) COI LCO gene primers

Primer	Sequence	Tm (°C)	GC (%)	Product size
Forward	5'-GGTCAACAAATCATAAAGATATTGG-3	55	54.5	658 base pair
Reverse	5'-TAAACTTCAGGGTGACCAAAAAATCA-3'	55	52.4	

Diagnosis of Gene

Figure (1) shows the results of the extracted DNA samples from the entire legs of the American cockroach, 17 samples from six different cities in Iraq. The first two samples were from the city of Samarra in central Iraq, three samples from the city of Baghdad in central Iraq, three samples from the city of Nasiriyah and Basra in southern Iraq, and three samples each from the cities of Kirkuk and Dohuk in northern Iraq. The samples showed the concentration of the extracted DNA (22.9, 6.8, 17.6, 7.9, 22.5, 11.4, 8.1, 19.5, 10.5, 13.6, 6.9, 11.8, 8.4, 7.3, 18.6, 6.0, 28.3). The samples were transferred on the electrophoresis device using agarose gel at a concentration of 1% and an electrical potential difference of 5 volts/cm² for an hour. The gel was then photographed under ultraviolet rays (UV). The results showed that the bands resulting from the transfer process using a high amount of purity, if the purity ranged for all samples From 1.6 to 1.8, this is the acceptable value for performing a PCR for the COX, COI LCO genes. Only cockroach legs were used to obtain the best results and

avoid the presence of proteins, fats, and sugars. Cockroach legs are characterized by low nutritional content, as well as hormones and digestive enzymes. They contain well-formed skeletal muscles, which facilitates the extraction process. The extracted samples were registered in the Global GenBank. The MTCOI gene is the most widely used gene in insect diagnosis because it has several different shapes and sizes. It can be used as a primary tool in differentiating between insect species, even within the same genus. If the difference between the American and German cockroaches is 25%, then the MTCOI gene is an important diagnostic tool in insects (Sharawis & Assagafa: 2021).

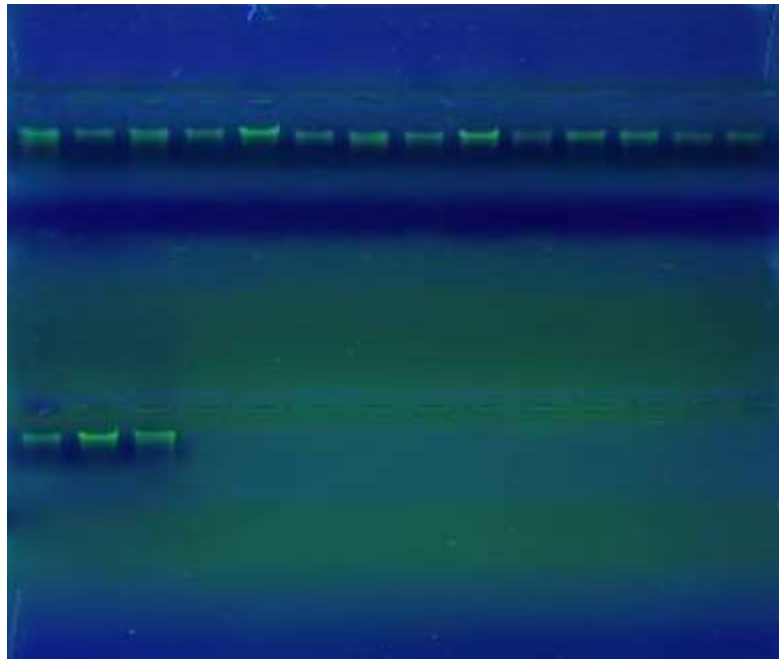


Figure (4) The result of DNA transfer

COX Gene Amplification

The results of the COX gene expression from mitochondrial DNA in the leg cells of the American cockroach were obtained from six samples collected from different cities in Iraq. PCR results were carried out using an electrophoresis device on a 1% agarose gel at a voltage of 5 V/cm² for an hour. The gel was then imaged under ultraviolet light. The results showed that the bands resulting from the PCR process, under the appropriate conditions for COX gene amplification, had a molecular weight of 420 base pairs, as shown in Figure (2). Gene 1. COX is the CYTCHROME C OXIDASE. Cytochrome C oxidase is the key enzyme in Complex IV of the electron transport chain (ETC) in mitochondria. It plays a crucial role in energy production within living cells(Yin et al,2022; Cox,2021; Wang et al,2023).

Especially in aerial organisms such as flying insects and humans, COX is a multi-component protein complex consisting of three subunits encoded by the nuclear genome of eukaryotes. It is the final enzyme for all aerobic metabolism. It plays an important role in energy balance and energy production in the form of ATP. It is found largely in muscles, particularly the pectoral muscles and leg muscles, which require high energy, particularly flight and jumping muscles. It is also abundant in insects with high resistance to pesticides, such as cockroaches and rice weevils. The biological function of COX is that this enzyme catalyzes the final reaction in the electron transport chain, catalyzing the transfer of electrons from the cytochrome C molecule to molecular oxygen (O₂), resulting in its reduction to water (H₂O)(Blomberg,2021; Noodleman et al,2023; Saura et al,2022)

It is one of the most widely used genes in insect classification and determining the genetic similarities and divergences between different species. COX also represents a major site for regulating oxidoreductase phosphorylation (OXPHS). The function of COX is to transfer electrons from cytochrome c to molecular oxygen, which leads to the formation of water and the release of energy needed to produce ATP. The enzyme cytochrome c oxidase (COX) is one of the most important tools in the study of genetic convergence and molecular evolution, especially through the analysis of the genes of the first subunit (COX1 or MT-CO1) located in mitochondrial DNA (mtDNA) (Mesquita et al., 2021; Kadenbach,2018; Ramzan et al ,2021).

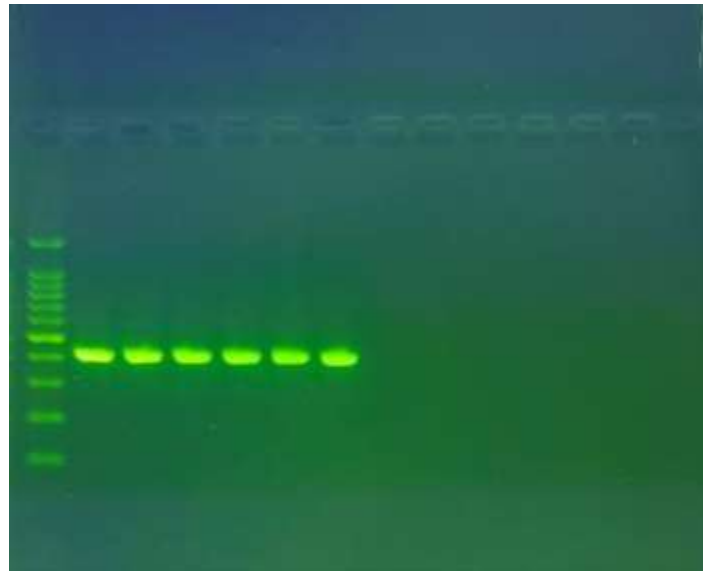


Figure (4) The result of gene transfer after performing PCR COX1.

Results of the COX1 gene sequencing

After amplifying the COX1 gene using the PCR technique, the sequences of the nitrogenous bases were determined for six samples from six different cities in Iraq. The PCR products were sent to a laboratory in South Korea.

BASRA 400
Query 1 ATATAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAG 60
|||||
Sbjct 249 ATATAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAG 308
Query 61 AAAGAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATG 120
|||||
Sbjct 309 AAAGAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATG 368
Query 121 CCGGAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTC 180
|||||
Sbjct 369 CCGGAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTC 428
Query 181 TAGGAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAG 240
|||||
Sbjct 429 TAGGAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAG 488
Query 241 AACGAATTCCTTTTCGTATGATCAGTAGCTATTACAGCattattatta ttattatCTC 300
|||||
Sbjct 489 AACGAATTCCTTTTCGTATGATCAGTAGCTATTACAGCATTATTATTATTATTATCTC 548
Query 301 TACCAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCT 360
|||||
Sbjct 549 TACCAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCT 608
Query 361 TTTTGATCCAGCAGGAGGGG 381
|||||
Sbjct 609 TTTTGATCCAGCAGGAGGGG 629

Nasiriyah 400
Query 1 ATATAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAG 60
|||||
Sbjct 249 ATATAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAG 308

Query 61 AAAGAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATG 120
|||||
Sbjct 309 AAAGAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATG 368
Query 121 CCGGAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTC 180
|||||
Sbjct 369 CCGGAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTC 428
Query 181 TAGGAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAG 240
|||||
Sbjct 429 TAGGAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAG 488
Query 241 AACGAATTCCCCTTTTCGTATGATCAGTAGCTATTACAGCATTATTATATTATTATCTC 300
|||||
Sbjct 489 AACGAATTCCCCTTTTCGTATGATCAGTAGCTATTACAGCATTATTATATTATTATCTC 548
Query 301 TACCAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCT 360
|||||
Sbjct 549 TACCAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCT 608
Query 361 TTTTGATCCAGCAGGAGGGGG 382
|||||
Sbjct 609 TTTTGATCCAGCAGGAGGGGG 630

Baghdad 379
Query 1 TAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAA 60
|||||
Sbjct 252 TAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAA 311
Query 61 GAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCG 120
|||||
Sbjct 312 GAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCG 371
Query 121 GAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAG 180
|||||
Sbjct 372 GAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAG 431
Query 181 GAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAAC 240
|||||
Sbjct 432 GAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAAC 491
Query 241 GAATTCCCCTTTTCGTATGATCAGTAGCTATTACAGCattattattattatCTCTAC 300
|||||
Sbjct 492 GAATTCCCCTTTTCGTATGATCAGTAGCTATTACAGCATTATTATTATTATTATCTCTAC 551

Query 301 CAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTT 360
|||||
Sbjct 552 CAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTT 611
Query 361 TTGATCCAGCAGGAGGGGG 379
|||||
Sbjct 612 TTGATCCAGCAGGAGGGGG 630

Samarra 400
Query 1 ATAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAA 60
|||||
Sbjct 251 ATAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAA 310

Query 61 AGAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCC 120
|||||
Sbjct 311 AGAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCC 370
Query 121 GGAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTA 180
|||||
Sbjct 371 GGAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTA 430
Query 181 GGAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAA 240
|||||
Sbjct 431 GGAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAA 490
Query 241 CGAATCCCCCTTTTCGTATGATCAGTAGCTATTACAGCattattattattattCTCTA 300
|||||
Sbjct 491 CGAATCCCCCTTTTCGTATGATCAGTAGCTATTACAGCATTATTATTATTATTATCTCTA 550
Query 301 CCAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTT 360
|||||
Sbjct 551 CCAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTT 610
Query 361 TTTGATCCAGCAGGAGGGGG 380
|||||

Sbjct 611 TTTGATCCAGCAGGAGGGGG 630

Kirkuk 397

Query 1 TAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAA 60
|||||
Sbjct 252 TAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAA 311
Query 61 GAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCG 120
|||||
Sbjct 312 GAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCG 371
Query 121 GAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAG 180
|||||
Sbjct 372 GAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAG 431
Query 181 GAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAAC 240
|||||
Sbjct 432 GAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAAC 491
Query 241 GAATCCCCCTTTTCGTATGATCAGTAGCTATTACAGCattattattattattCTCTAC 300
|||||
Sbjct 492 GAATCCCCCTTTTCGTATGATCAGTAGCTATTACAGCATTATTATTATTATTATCTCTAC 551
Query 301 CAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTT 360
|||||
Sbjct 552 CAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTT 611
Query 361 TTGATCCAGCAGGAGGGGG 379
|||||

Sbjct 612 TTGATCCAGCAGGAGGGGG 630

DHUK 405

Query 7 ATAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAA 66
|||||
Sbjct 251 ATAAGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAA 310
Query 67 AGAGGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCC 126

Sbjct 311 AGAGGTGCCGGAACAGGATGAACAGTATACCCACCCTAGCAAGAGGCATTGCTCATGCC 370
Query 127 GGAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTA 186
|||||
Sbjct 371 GGAGCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTA 430
Query 187 GGAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAA 246
|||||
Sbjct 431 GGAGCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAA 490
Query 247 CGAATTCCTCTTTTCGTATGATCAGTAGCTATTACAGCattattattattattCTCTA 306
|||||
Sbjct 491 CGAATTCCTCTTTTCGTATGATCAGTAGCTATTACAGCATTATTATTATTATTATCTCTA 550
Query 307 CCAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTT 366
|||||
Sbjct 551 CCAGTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTT 610
Query 367 TTTGATCCAGCAGGAGGGGG 386
|||||
Sbjct 611 TTTGATCCAGCAGGAGGGGG 630

2. THE AMERICAN COCKROACH PHYLOGENETIC TREE FOR THE COX GENE

Figure (3) The genetic tree of the American cockroach for the COX1 gene. Using the National Center for Biotechnology Information (NCBI) website, then accessing the BLAS NUCLEOTIDE subwindow and comparing them to the recorded species, the species were 100% similar to each other, 100% identical to the recorded species NC016956.1 from the United States, 100% identical to the recorded species OL589367 from Vietnam, 67% identical to the recorded species 220409.1 from Malaysia, and 93% identical to the recorded species LC619069.1 from Japan.

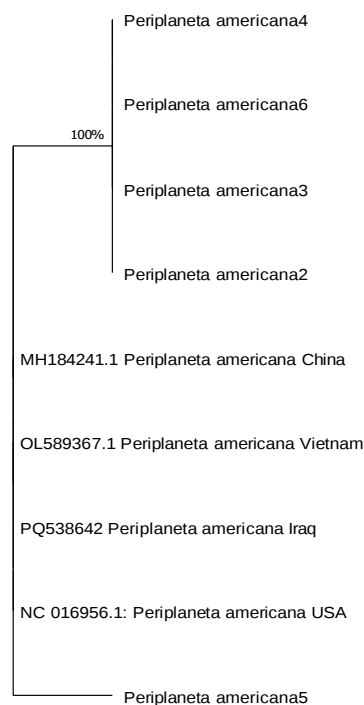


Figure (3) The genetic tree of the American cockroach for the COX1 gene

3. RESULTS OF THE COI LC GENE MIGRATION

The results of the COI LC gene migration from mitochondrial DNA in the leg cells of the American cockroach were shown for six samples collected from different cities in Iraq. PCR results were migration using an electrophoresis device on agarose gel at a concentration of 2% and an electrical potential difference of 5 V/cm² for an hour, then the gel was imaged under ultraviolet light.

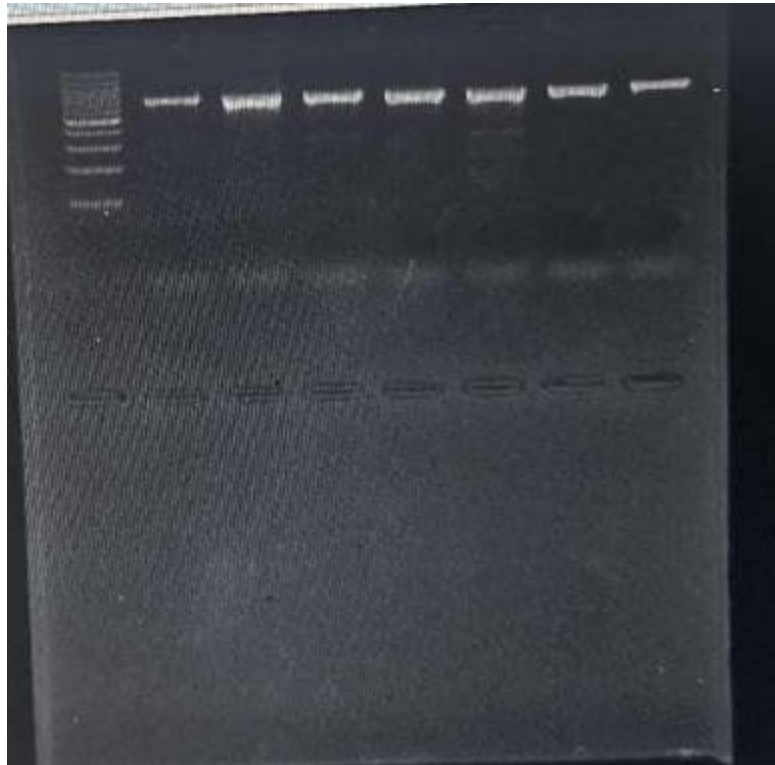


Figure (4) The result of gene transfer after performing PCR COI LC.

Then the results showed that the bands resulting from the migration process using the PCR technique and according to the appropriate conditions for the COI LCO gene assembly had a molecular weight of 580 base pairs as shown in Figure (4-5).

4. RESULTS OF THE COI LC GENE SEQUENCING

After amplifying the COI LC gene using the PCR technique, the sequences of the nitrogenous bases were determined for six samples from six different cities in Iraq. The PCR products were sent to an external laboratory, and the results were presented.

BASRA

```
Query 15  ATTTTCGGTGCCTGATCAGGTATAGTGGGAACATCACTAAGAATATTAATTCGTGCTGAG 74
          ||||||||||||||||||||||||||||||||||||||||||||||||||||||||||
Sbjct 14  ATTTTCGGTGCCTGATCAGGTATAGTGGGAACATCACTAAGAATATTAATTCGTGCTGAG 73
Query 75  CTCGGGCAGCCAGGTTCACTAATTGGAGATGATCAAATTTATAATGTAATTGTTACTGCC 134
          ||||| |||||||||||||||||||||||||||||
Sbjct 74  CTCGGGCAACCAGGTTCACTAATTGGAGATGATCAAATTTATAATGTAATCGTTACTGCC 133
Query 135 CATGCTTTCATTATAATTTTCTTTATAGTAATACCAATCATAATTGGGGGATTTGGTAAT 194
          ||||| |||||||||||||||||||||||||||||
Sbjct 134 CATGCCTTCATTATAATTTTCTTTATAGTAATACCAATCATAATTGGGGGATTTGGTAAT 193
Query 195 TGATTAGTACCACTAATATTAGGAGCCCCAGATATAGCCTTCCCACGAATAAATAATATA 254
          ||||||||||||||||||||||||||||||||||
Sbjct 194 TGATTAGTACCACTAATATTAGGAGCCCCAGATATAGCCTTCCCACGAATAAATAATATA 253
Query 255 AGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTAGTAGAAAGA 314
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|||||
Sbjct 254 AGATTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAAGA 313
Query 315 GGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGGA 374
|||||
Sbjct 314 GGTGCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGGA 373
Query 375 GCATCTGTTGATCTAGCAATTTTTCTTTACATCTAGCAGGTGTATCCTCAATTCTAGGA 434
|||||
Sbjct 374 GCATCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAGGA 433
Query 435 GCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAACGA 494
|||||
Sbjct 434 GCTGTAAATTTTATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAACGA 493
Query 495 ATTCCCCTTTTCGTATGATCAGTAGCTATTACAGCattattattattatCTCTACCA 554
|||||
Sbjct 494 ATTCCCCTTTTCGTATGATCAGTAGCTATTACAGCATTATTATTATTATTATCTCTACCA 553
Query 555 GTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTTTT 614
|||||
Sbjct 554 GTGCTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTTTT 613
Query 615 GATCCAGCAGGAGGGGGTGATCCAATTTTATATCAACACTTATTC 659
|||||
Sbjct 614 GATCCAGCAGGAGGGGGTGACCCAATTTTATATCAACACTTATTC 658

Nasiriyah

Query 34 AGCTATAGTGGGAACATCACTAAGAATATTAATTCGTGCTGAGCTCTGGGCAACCAGGTT 93 ||
|||||
Sbjct 31 AGGTATAGTGGGAACATCACTAAGAATATTAATTCGTGCTGAGCTC-GGGCAACCAGGTT 89
Query 94 CATTAATTGGAGATGATCAAATTTATAATGTAATCGTTACTGCCCATGCTTTCATTATAA 153
|| |||||
Sbjct 90 CACTAATTGGAGATGATCAAATTTATAATGTAATCGTTACTGCCCATGCCTTCATTATAA 149
Query 154 TTTCTTTATAGTTATACCAATCATAATTGGGGGATTTGGTAATTGATTAGTACCACTAA 213
|||||
Sbjct 150 TTTCTTTATAGTAATACCAATCATAATTGGGGGATTTGGTAATTGATTAGTACCACTAA 209
Query 214 TATTAGGAGCCCCAGATATAGCCTTCCCACGAATAAATAACATAAGATTCTGATTATTAC 273
|||||
Sbjct 210 TATTAGGAGCCCCAGATATAGCCTTCCCACGAATAAATAATATAAGATTCTGATTATTAC 269
Query 274 CACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAAGAGGTGCCGGAACAGGAT 333
|||||
Sbjct 270 CACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAAGAGGTGCCGGAACAGGAT 329
Query 334 GAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGAGCATCTGTTGATCTAG 393
|||||
Sbjct 330 GAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGAGCATCTGTTGATCTAG 389
Query 394 CAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAGGAGCTGTAAATTTTATCT 453
|||||
Sbjct 390 CAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAGGAGCTGTAAATTTTATCT 449
Query 454 CCACAACAATTAATATAAAACCTATTAATATAAAACCAGAACGAATTCCCCTTTTCGTAT 513
|||||
Sbjct 450 CCACAACAATTAATATAAAACCTATTAATATAAAACCAGAACGAATTCCCCTTTTCGTAT 509

Query 514 GATCAGTAGCTATTACAGCattattattattatCTCTACCAGTGCTTGCTGGAGCAA 573

|||||

Sbjct 510 GATCAGTAGCTATTACAGCATTATTATTATTATTATCTCTACCAGTGCTTGCTGGAGCAA 569

Query 574 TTAATAATTAACTGACCGAAATCTAAATACATCCTTTTTTGATCCAGCAGGAGGGG 633

|||||

Sbjct 570 TTAATAATTAACTGACCGAAATCTAAATACATCCTTTTTTGATCCAGCAGGAGGGG 629

Query 634 GTGACCCAATTTTATATCAACACTTATTC 662

|||||

Sbjct 630 GTGACCCAATTTTATATCAACACTTATTC 658

Baghdad

Query 17 TTCGGTG-CTGATCAGGCATAGTGGAACATCACTAAGAATATTAATTCGTGCTGAGCTC 75

|||||

Sbjct 17 TTCGGTGCTGATCAGGTATAGTGGAACATCACTAAGAATATTAATTCGTGCTGAGCTC 76

Query 76 GGGCAACCAGGTTTCTTAATTGGAGATGATCAAATTTATAATGTAATCGTTACTGCCCAT 135

|||||

Sbjct 77 GGGCAACCAGGTTTCTTAATTGGAGATGATCAAATTTATAATGTAATCGTTACTGCCCAT 136

Query 136 GCTTTCATTATAATTTTCTTTATAGTTATACCAATCATAATTGGGGGATTTGGTAATTGA 195

|||||

Sbjct 137 GCCTTCATTATAATTTTCTTTATAGTAATACCAATCATAATTGGGGGATTTGGTAATTGA 196

Query 196 TTAGTACCACTAATATTAGGAGCCCCAGATATAGCCTTCCCACGAATAAATAACATAAGA 255

|||||

Sbjct 197 TTAGTACCACTAATATTAGGAGCCCCAGATATAGCCTTCCCACGAATAAATAATATAAGA 256

Query 256 TTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAAGAGGT 315

|||||

Sbjct 257 TTCTGATTATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAAGAGGT 316

Query 316 GCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGGAGCA 375

|||||

Sbjct 317 GCCGGAACAGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGGAGCA 376

Query 376 TCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAGGAGCT 435

|||||

Sbjct 377 TCTGTTGATCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAGGAGCT 436

Query 436 GTAAATTTTATCTCCACAACAATTAATATAAAAACCTATTAATATAAAAACCAGAACGAATT 495

|||||

Sbjct 437 GTAAATTTTATCTCCACAACAATTAATATAAAAACCTATTAATATAAAAACCAGAACGAATT 496

Query 496 CCCCTTTTCGTATGATCAGTAGCTATTACAGCattattattattatCTCTACCAGTG 555

|||||

Sbjct 497 CCCCTTTTCGTATGATCAGTAGCTATTACAGCATTATTATTATTATTATCTCTACCAGTG 556

Query 556 CTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTTTTGAT 615

|||||

Sbjct 557 CTTGCTGGAGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTTTTGAT 616

Query 616 CCAGCAGGAGGGGGTGACCCAATTTTATATCAACACTTATTC 657

|||||

Sbjct 617 CCAGCAGGAGGGGGTGACCCAATTTTATATCAACACTTATTC 658

Samarra

Query 1 ATAGTGGAACATCACTAAGAATATTAATTCGTGCTGAGCTCGGGCAACCAGGTTTCTTA 60

|||||
Sbjct 35 ATAGTGGGAACATCACTAAGAATATTAATTCGTGCTGAGCTCGGGCAACCAGGTTCACTA 94
Query 61 ATTGGAGATGATCAAATTTATAATGTAATCGTTACTGCCCATGCTTTCATTATAATTTTC 120
|||||
Sbjct 95 ATTGGAGATGATCAAATTTATAATGTAATCGTTACTGCCCATGCCTTCATTATAATTTTC 154
Query 121 TTTATAGTTATACCAATCATAATTGGGGGATTTGGTAATTGATTAGTACCACTAATATTA 180
|||||
Sbjct 155 TTTATAGTAATACCAATCATAATTGGGGGATTTGGTAATTGATTAGTACCACTAATATTA 214
Query 181 GGAGCCCCAGATATAGCCTTCCCACGAATAAATAACATAAGATTCTGATTATTACCACCT 240
|||||
Sbjct 215 GGAGCCCCAGATATAGCCTTCCCACGAATAAATAATATAAGATTCTGATTATTACCACCT 274
Query 241 TCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAAGAGGTGCCGGAACAGGATGAACA 300
|||||
Sbjct 275 TCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAAGAGGTGCCGGAACAGGATGAACA 334
Query 301 GTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGAGCATCTGTTGATCTAGCAATT 360
|||||
Sbjct 335 GTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGAGCATCTGTTGATCTAGCAATT 394
Query 361 TTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAGGAGCTGTAAATTTTATCTCCACA 420
|||||
Sbjct 395 TTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAGGAGCTGTAAATTTTATCTCCACA 454
Query 421 ACAATTAATATAAAAACCTATTAATATAAAAACCAGAACGAATTCCCCTTTTCGTATGATCA 480
|||||
Sbjct 455 ACAATTAATATAAAAACCTATTAATATAAAAACCAGAACGAATTCCCCTTTTCGTATGATCA 514
Query 481 GTAGCTATTACAGCattattattattatCTCTACCAGTGCTTGCTGGAGCAATTACT 540
|||||
Sbjct 515 GTAGCTATTACAGCATTATTATTATTATTATCTCTACCAGTGCTTGCTGGAGCAATTACT 574
Query 541 ATATTATTAAGTACCGAAATCTAAATACATCCTTTTTTGATCCAGCAGGAGGGGGTGAC 600
|||||
Sbjct 575 ATATTATTAAGTACCGAAATCTAAATACATCCTTTTTTGATCCAGCAGGAGGGGGTGAC 634
Query 601 CCAATTTTATATCAACACTTATTC 624
|||||
Sbjct 635 CCAATTTTATATCAACACTTATTC 658

KIRKUK

Query 1 CATCACTAAGAATATTAATTCGTGCTGAGCTCGGGCAACCAGGTTCAATTAATTGGAGATG 60
|||||
Sbjct 45 CATCACTAAGAATATTAATTCGTGCTGAGCTCGGGCAACCAGGTTCACTAATTGGAGATG 104
Query 61 ATCAAATTTATAATGTAATCGTTACTGCCCATGCTTTCATTATAATTTCTTTATAGTTA 120
|||||
Sbjct 105 ATCAAATTTATAATGTAATCGTTACTGCCCATGCCTTCATTATAATTTCTTTATAGTAA 164
Query 121 TACCAATCATAATTGGGGGATTTGGTAATTGATTAGTACCACTAATATTAGGAGCCCCAG 180
|||||
Sbjct 165 TACCAATCATAATTGGGGGATTTGGTAATTGATTAGTACCACTAATATTAGGAGCCCCAG 224
Query 181 ATATAGCCTTCCCACGAATAAATAACATAAGATTCTGATTATTACCACCTTCATTAACCTT 240
|||||

Sbjct 225 ATATAGCCTTCCCACGAATAAATAATATAAGATTCTGATTATTACCACCTTCATTAACCTT 284
Query 241 TATTACTAGCTAGTAGTATAGTAGAAAGAGGTGCCGGAACAGGATGAACAGTATACCCAC 300
|||||

Sbjct 285 TATTACTAGCTAGTAGTATAGTAGAAAGAGGTGCCGGAACAGGATGAACAGTATACCCAC 344
Query 301 CACTAGCAAGAGGCATTGCTCATGCCGGAGCATCTGTTGATCTAGCAATTTTTTCATTAC 360
|||||

Sbjct 345 CACTAGCAAGAGGCATTGCTCATGCCGGAGCATCTGTTGATCTAGCAATTTTTTCATTAC 404
Query 361 ATCTAGCAGGTGTATCCTCAATTCTAGGAGCTGTAAATTTTATCTCCACAACAATTAATA 420
|||||

Sbjct 405 ATCTAGCAGGTGTATCCTCAATTCTAGGAGCTGTAAATTTTATCTCCACAACAATTAATA 464
Query 421 TAAAACCTATTAATATAAAACCAGAACGAATTCCCCTTTTCGTATGATCAGTAGCTATTA 480
|||||

Sbjct 465 TAAAACCTATTAATATAAAACCAGAACGAATTCCCCTTTTCGTATGATCAGTAGCTATTA 524
Query 481 CAGCattattattattatCTCTACCAGTGCTTGCTGGAGCAATTACTATATTATTAA 540
|||||

Sbjct 525 CAGCATTATTATTATTATTATCTCTACCAGTGCTTGCTGGAGCAATTACTATATTATTAA 584
Query 541 CTGACCGAAATCTAAATACATCCTTTTTTGATCCAGCAGGAGGGGGTGACCCAATTTTAT 600
|||||

Sbjct 585 CTGACCGAAATCTAAATACATCCTTTTTTGATCCAGCAGGAGGGGGTGACCCAATTTTAT 644
Query 601 ATCAACACTTATTC 614
|||||

Sbjct 645 ATCAACACTTATTC 658

DHUK
TGATCAGAGTATAGTGGAACATCACTAAGAATATTAATTCGTGCTGAGCTCGGGCAACC 84
|||||

Sbjct 26 TGATCAG-GTATAGTGGAACATCACTAAGAATATTAATTCGTGCTGAGCTCGGGCAACC 84
Query 85 AGGTTCACTAATTGGAGATGATCAAATTTATAATGTAATCGTTACTGCCCATGCCTTCAT 144
|||||

Sbjct 85 AGGTTCACTAATTGGAGATGATCAAATTTATAATGTAATCGTTACTGCCCATGCCTTCAT 144
Query 145 TATAATTTTCTTTATAGTAATACCAATCATAATTGGGGGATTTGGTAATTGATTAGTACC 204
|||||

Sbjct 145 TATAATTTTCTTTATAGTAATACCAATCATAATTGGGGGATTTGGTAATTGATTAGTACC 204
Query 205 ACTAATATTAGGAGCCCCAGATATAGCCTTCCCACGAATAAATAATATAAGATTCTGATT 264
|||||

Sbjct 205 ACTAATATTAGGAGCCCCAGATATAGCCTTCCCACGAATAAATAATATAAGATTCTGATT 264
Query 265 ATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAAGAGGTGCCGGAAC 324
|||||

Sbjct 265 ATTACCACCTTCATTAACCTTTATTACTAGCTAGTAGTATAGTAGAAAGAGGTGCCGGAAC 324
Query 325 AGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGGAGCATCTGTTGA 384
|||||

Sbjct 325 AGGATGAACAGTATACCCACCACTAGCAAGAGGCATTGCTCATGCCGGAGCATCTGTTGA 384
Query 385 TCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAGGAGCTGTAAATTT 444
|||||

Sbjct 385 TCTAGCAATTTTTTCATTACATCTAGCAGGTGTATCCTCAATTCTAGGAGCTGTAAATTT 444

Query 445 TATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAACGAATCCCTTTT 504

|||||

Sbjct 445 TATCTCCACAACAATTAATATAAAACCTATTAATATAAAACCAGAACGAATCCCTTTT 504

Query 505 CGTATGATCAGTAGCTATTACAGCattattattattatCTCTACCAGTGCTTGCTGG 564

|||||

Sbjct 505 CGTATGATCAGTAGCTATTACAGCATTATTATTATTATCTCTACCAGTGCTTGCTGG 564

Query 565 AGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTTTTGATCCAGCAGG 624

|||||

Sbjct 565 AGCAATTACTATATTATTAAGTACCGAAATCTAAATACATCCTTTTTTGATCCAGCAGG 624

Query 625 AGGGGGTGACCCAATTTTATATCAACACTTATTC 658

|||||

Sbjct 625 AGGGGGTGACCCAATTTTATATCAACACTTATTC 658

Analysis of the Scansink results for the COILC gene

Tebil(1) shows the occurrence of many changes in the DNA strand through the presence of gaps, substitutions, inversions, and deletions in different ways for each sample.

Periplaneta Americana						
No.	Type of substitution	Location	Nucleotide	Sequence ID with compare	Source	Identities
1	GAP	33	A\-	.1	Periplaneta americana isolate NH20 cytochrome oxidase subunit I (COI) gene, partial cds; mitochondrial	99%
2	Transversion	33	C\G	: .1	Periplaneta americana isolate NH20 cytochrome oxidase subunit I (COI) gene, partial cds; mitochondrial	99%
	GAP	76	T\-			
	Transition	93	T\C			
	Transition	139	T\C			
	Transversion	164	T\A			
	Transversion	250	C\T			
3	GAP	24	-C	.1	Periplaneta americana isolate NH20 cytochrome oxidase subunit I (COI) gene, partial cds; mitochondrial	99%
	Transition	34	C\T			
	Transition	93	T\C			
	Transition	139	T\C			
	Transversion	163	T\A			
	Transition	250	C\T			
4	Transition	92	T\C	.1	Periplaneta americana isolate NH20 cytochrome oxidase subunit I (COI) gene, partial cds; mitochondrial	99%
	Transition	140	T\C			
	Transversion	163	T\A			
	Transition	250	C\T			
5	Transition	82	G\A	.1	Periplaneta americana isolate NH20 cytochrome oxidase subunit I (COI)	99%
	Transition	126	T\C			

	Transi tion	139	TAC		gene, partial cds; mitochondrial	
	Transversion	400	TAA			
	Transition	635	TAC			
6	Transition	92	TAC	.1	Periplaneta americana isolate NH20 cytochrome oxidase subunit I (COI) gene, partial cds; mitochondrial	99%
	Transition	139	TAC			
	Transversion	163	TAA			
	Transition	250	CAT			

Table (1) shows the differences between the studied samples.

The American cockroach phylogenetic tree for the COILC gene

After accessing the GenBank website, going to the BLAST page and searching for the sequence for each sample in the search, the search results showed:

The results showed that the sample isolated from Basra matched the sample bearing the global number MH184229.1 with a 97% match rate, which is from China

The results showed that the sample isolated from Nasiriyah matched the sample bearing the global number MF149291.1 with a 97% match rate, which is from China.

The results showed that the sample isolated from Baghdad matched the sample bearing the global number MH1840721.1 with a 99% match rate, which is from Thailand.

The results showed that the sample isolated from Samarra matched the sample bearing the global number MH184075.1 with a 95% match rate, which is from China. The results showed that the sample isolated from Kirkuk city matched the sample bearing the global number MH184072.1 with a 97% match rate, which is from China.

The results showed that the sample isolated from Dohuk city matched the sample bearing the global number MF149272.1 with a 97% match rate, which is from China.

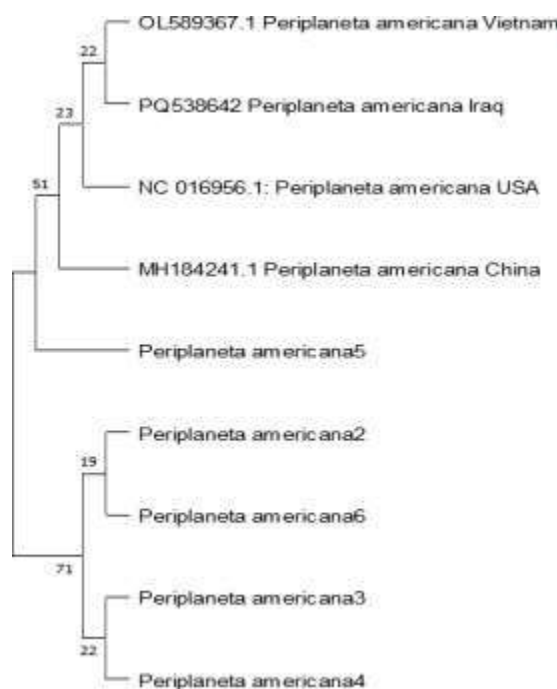


Figure (5) The genetic tree of the American cockroach for the COILC gene.

REFERENCES

- [1] Ademolu, K. O., Mustapha, O. Y., & Idowu, A. B. (2020). Nutritional and gut microbial analyses of adult male cockroaches (*Periplaneta americana*)(Dictyoptera, Blattodea) from three locations in Abeokuta, Nigeria. *Entomologica romanica*, 24, 19-23.
 - [2] Cerreta, A. J., Smith, D. C., Ange-Van Heugten, K., & Minter, L. J. (2022). Comparative nutrient analysis of four species of cockroaches used as food for insectivores by life stage, species, and sex. *Zoo Biology*, 41(1), 26-33.
 - [3] Dominic, M. I. S., & Ab Majid, A. H. (2023). The draft genome dataset of the American cockroach, *Periplaneta americana* (Linnaeus, 1758)(Blattidae: Blattinae). *Data in Brief*, 49, 109301.
 - [4] Hamanaka, Y., Minoura, R., Nishino, H., Miura, T., Mizunami, M., 2016. Dopamine- and Tyrosine Hydroxylase-Immunoreactive Neurons in the Brain of the American Cockroach, *Periplaneta americana*. *PLoS One* 11 (8), e0160531. [https://doi.org/ 10.1371/journal.pone.0160531](https://doi.org/10.1371/journal.pone.0160531).
 - [5] Blomberg, M. R. (2021). The redox-active tyrosine is essential for proton pumping in cytochrome c oxidase. *Frontiers in chemistry*, 9, 640155.
 - [6] Noodleman, L., Götz, A. W., Han Du, W. G., & Hunsicker-Wang, L. (2023). Reaction pathways, proton transfer, and proton pumping in ba3 class cytochrome c oxidase: Perspectives from DFT quantum chemistry and molecular dynamics. *Frontiers in Chemistry*, 11, 1186022.
 - [7] Saura, P., Riepl, D., Frey, D. M., Wikström, M., & Kaila, V. R. (2022). Electric fields control water-gated proton transfer in cytochrome c oxidase. *Proceedings of the National Academy of Sciences*, 119(38), e2207761119.
 - [8] Cox, M. M. (2021). Innovations in the insect cell expression system for industrial recombinant vaccine antigen production. *Vaccines*, 9(12), 1504.
 - [9] Wang, L., Lai, Y., Chen, J., Cao, X., Zheng, W., Dong, L., & Wang, S. (2023). The ASH1 –PEX16 regulatory pathway controls peroxisome biogenesis for appressorium-mediated insect infection by a fungal pathogen. *Proceedings of the National Academy of Sciences*, 120(4), e2217145120.
 - [10] Kadenbach, B. (2018). Regulation of mitochondrial respiration and ATP synthesis via cytochrome c oxidase. *Rendiconti Lincei. Scienze Fisiche e Naturali*, 29, 421-435.
 - [11] Ramzan, R., Kadenbach, B., & Vogt, S. (2021). Multiple mechanisms regulate eukaryotic cytochrome c oxidase. *Cells*, 10(3), 514.
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