

## Isolation and identification of *Acetobacter* from Toddy sample for production of Acetic Acid

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### ABSTRACT

This study was concerned with the isolation and characterization of *Acetobacter aceti* from toddy for acetic acid production. The samples were inoculated in sterilized GYP standard media and then incubated at 28°C for 48 hours. Successive subculture was performed to screen out the strains. In Gram's staining, the morphology of the isolated bacteria exhibited pink, small rod shaped single, pair and chain in arrangement, in the hanging drops technique, all the isolates revealed motile. Biochemical tests were performed by fermentation of five basic sugars by producing both acid and gas bubbles in Durham tube. Acidity of the media gradually increased from day 0 to day 8 and pH of the media decreased from 6.8 to 5.5 during the period. This protocol was successful for enriching *Acetobacter aceti*, which was essential for Acetic acid production.

**Keywords:** Toddy sample Acetic acid, *Acetobacter* spp.

### 1. INTRODUCTION

Manzi is a typical tropical alcoholic beverage, produced by fermentation of sugary coconut sap is trapped from palm tree that grow at the coastal region of maharashtra. manzi is a white-shish effervescent,acidic alcoholic beverage (Swins and De Ley,1977) trapping process of toddy used in this study was as explained by Kadare *et al.* (2004).it is a product of mixed alcoholic lactic and acetic fermentation. AAB (acetic acid bacteria utilizing the glucose and sucrose might be present in an earlier stage of the manzi fermentation (okafar,1975) ***Acetobacter*** is a genus of acetic acid bacteria. Acetic acid bacteria are characterized by the ability to convert ethanol to acetic acid in the presence of oxygen. Of these, the genus *Acetobacter* is distinguished by the ability to oxidize lactate and acetate into carbon dioxide and water. Bacteria of the genus *Acetobacter* have been isolated from industrial vinegar fermentation processes and are frequently used as fermentation starter cultures. Acetic acid bacteria (AAB) is a group of gram-negative, aerobic and motile rods that carry out incomplete oxidation of alcohol and sugars, leading to the accumulation of organic acids as end products. These bacteria are heterogeneous assemblage organisms (Sokollek *et al.*, 1998; Madigan *et al.*, 2008). Among the most important acetic acid bacteria, the strains of genus *Acetobacter* are mainly involved in vinegar production. *Acetobacter* is a gram negative, obligate aerobe coccus or rod-shaped bacterium with the size of 0.6-0.8 X 1.0 - 4.0 µm, nonmotile or motile with peritrichous flagella, catalase positive and oxidase negative biochemically.

### 2. MATERIAL & METHODS

The coconut wine or toddy were collected 4 sample for Acetic acid production were obtained from Patan coastal region of Maharashtra .The freshly trapped toddy samples were collected in sterile cantainer.

### Isolation of *Acetobacter* spp. from toddy sample.

#### 1)Sample collection:

Toddy sample was collected from Patan Tal-Karad Dist.-Satara of coastal region of Maharashtra. Sample was collected in sterile container Daniel *et al.*2021).

Toddy sample was enriched by GYP Broth In 1week incubation period at a 30°C room temperature. ( Daniel *et al.*2021).

#### 3)Isolation method:

Isolation of microorganisms from Toddy .it was along enriched sample and spread on GYP Agar plates Agar plates and incubate at room temperature for 2 to 3 days. ( W Scharf *et al.* 2013).

#### 4)Chemical Analysis Determination of Acidity:

The method of SLS: 168:1999 was used. The acidity of the sample was determined as acetic acid (% w/w) by titrating 10 ml of the sample against 0.1 N NaOH using phenolphthalein as an indicator.

#### 5)Production of acetic acid using the submerged fermentation process

The effect of temperature (28, 30 and 32 °C), shaking (200 rpm) and inoculum size (5, 10, 20 and 30 ml) on formation of acetic acid in submerged fermentation were studied.

#### 6)Confirmation Test of Acetic Acid.

When the litmus test is done, acetic acid turn blue litmus to red. Acetic acid reacts with bases to form salt of sodium acetate and water. its acid -base neutralization reaction.

#### 7)Estimation of Acetic Acid

The production medium is aerated and samples were taken at 48 hrs interval .Estimation of acetic acid -5 m L of cultured is mix with 20 m L. Distilled water .3-5 drop phenolphthalein indicator is add .the solution is titrated against 0.5 N NaOH the amount of acetic acid produce in 100 ml of medium is calculate using the titration reading  $\times 0.03 \times 20$  (Guillaume-François Rouelle *et al.*1754).

Acetic acid (g/100ml)=

Volume of 0.5N NaOH(ml)used in titration $\times 0.03 \times 20$

### 3. RESULT

After incubation to check the staining ,biochemical and morphological characterization.and identified as AAB according to the standard guidelines of Bergey's manual of Determinative Bacteriology



Pure cultures of AAB isolates

variable +- positive – negative

Biochemical characterization of AAB

### Biochemical Characterization

4 isolates showing significant zones of clearance on GYP agar and which were found to be catalase positive and oxidase negative were preliminarily identified as AAB according to the standard guidelines of Bergey's manual of Determinative Bacteriology

### Chemical Analysis Determination of Acidity:

Acidity obtained from the medium having toddy inoculated with A1, A2 and control (without bacterial strain) was significantly increased

### Production of acetic acid using the submerged fermentation process

There were no significant differences in the formation of acetic acid (8.74%) between the size of the inoculum, whereas, that of acetic acid formation showed a significant difference during the fermentation period. Inoculum's size, such as 5, 10 and 20 % showed the same acetic acid production A3 (4.5%) at 8th, 5th and 7th day of fermentation respectively

### Estimation of Acetic Acid

BACTERIAL STARIN	A1	A2	A3	A4
Gram Stain	V	V	V	+
Catalase	+	+	+	+
Oxidase	-	-	-	-
Growth In Gelatine	+	+	+	+
Growth In pH2.5	-	-	-	-
Growth In pH 4.5	+	+	+	+
Growth In pH 7	+	+	+	+
Motility	+	+	+	+

Sr.no	Sample	Total yield
1	A1	3.5%
2	A2	4.2%
3	A3	4.5%
4	A4	2.3%

### 4. CONCLUSION

In this study, acetic acid-producing bacteria were selected by using toddy , based on the acetic acid production, morphological characterization also the formation of gel-like layers in the fermentation medium. Among the three processes of acetic acid production, the submerged process yielded the highest amount of acetic acid A3 (4.5%)at the 8<sup>th</sup> day of fermentation

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### Competing interest

There is high potential Competing interest

## REFERENCES

- [1] Andrés-Barrao C, Benagli C, Chappuis M, Ortega Pérez R, Tonolla M, Barja F. Rapid identification of acetic acid bacteria using MALDI-TOF mass spectrometry fingerprinting. *Syst Appl Microbial*. 2013;36(2):75–81. 10.1016/j.syapm.2012.09.002.
- [2] Asai T. Taxonomic studies on acetic acid bacteria and allied oxidative bacteria isolated from fruits: A new classification of the oxidative bacteria. *Nippon Nogeikagaku Kaishi*. 1934;10:621–9. [in Japanese] 10.1271/nogeikagaku1924.10.621
- [3] Asai T. Taxonomic studies on acetic acid bacteria and allied oxidative bacteria isolated from fruits: A new classification of the oxidative bacteria. *Nippon Nogeikagaku Kaishi*. 1935;11:674–708. [in Japanese] 10.1271/nogeikagaku1924.11.8\_674
- [4] Beijerinck MW. About the types of acetic acid bacteria. *Zentralbl Bakteriol Parasitenkd Infektionskr Hyg Abt II*. 1898;4:209–16. [in German]
- [5] Buchanan RE, Gibbons NE, editors. *Bergey's manual of determinative bacteriology*. Baltimore, MD, USA: Williams & Wilkins Co; 1974.
- [6] Cacicedo ML, Castro MC, Servetas I, Bosnea L, Boura K, Tsafrakidou P, et al. Progress in bacterial cellulose matrices for biotechnological applications. *Bioresour Technol*. 2016;213:172–80. 10.1016/j.biortech.2016.02.071
- [7] Cleenwerck I, De Vos P. Polyphasic taxonomy of acetic acid bacteria: An overview of the currently applied methodology. *Int J Food Microbiol*. 2008;125(1):2–14. 10.1016/j.ijfoodmicro.2007.04.017
- [8] De Oliveira ALD, Santos V, Junior, Liotti RG, Zilioli E, Spinosa WA, Ribeiro-Paes JT. Study of bacteria *Gluconobacter* sp.: Isolation, purification, phenotypic and molecular identification. *Food Sci Technol (Campinas)*. 2010;30(1):106–12. 10.1590/S0101-20612010000100016
- [9] Dikshit PK, Moholkar VS. Optimization of 1,3-dihydroxyacetone production from crude glycerol by immobilized *Gluconobacter oxydans* MTCC 904. *Bioresour Technol*. 2016;216:1058–65. 10.1016/j.biortech.2016.01.100
- [10] Ebner H, Follmann H. Acetic Acid. In: Rehm HJ, Reed G, editors. *Biotechnology*. Weinheim, Germany: Wiley-VCH; 1983. pp. 389–407.
- [11] Fernández-Pérez R, Torres C, Sanz S, Ruiz-Larrea F. Rapid molecular methods for enumeration and taxonomical identification of acetic acid bacteria responsible for submerged vinegar production. *Eur Food Res Technol*. 2010;231(5):813–9. 10.1007/s00217-010-1331-6
- [12] Frateur J. Essay on the systematics of *Acetobacters*. *Cellule*. 1950;53:287–392. [in French]
- [13] Hansen EC. Research on acidifying bacteria. *C R Trav Lab Carlsberg*. 1894;3:182–216. [in French]
- [14] Hattori H, Yakushi T, Matsutani M, Moonmangmee D, Toyama H, Adachi O, et al. High-temperature sorbose fermentation with thermotolerant *Gluconobacter frateurii* CHM43 and its mutant strain adapted to higher temperature. *Appl Microbiol Biotechnol*. 2012;95(6):1531–40. 10.1007/s00253-012-4005-4
- [15] Hu ZC, Liu ZQ, Zheng YG, Shen YC. Production of 1,3-dihydroxyacetone from glycerol by *Gluconobacter oxydans* ZJB09112. *J Microbiol Biotechnol*. 2010;20(2):340–5.
- [16] *Int J Food Microbiol*. 2015;196:137–44. 10.1016/j.ijfoodmicro.2014.12.003
- [17] Nakano S, Fukaya M. Analysis of proteins responsive to acetic acid in *Acetobacter*: Molecular mechanisms conferring acetic acid resistance in acetic acid bacteria. *Int J Food Microbiol*. 2008;125(1):54–9. 10.1016/j.ijfoodmicro.2007.05.015
- [18] Spinosa WA. Isolation, selection, identification and kinetic parameters of acetic acid bacteria from vinegar industry [PhD Thesis]. Campinas, Brazil: State University of Campinas; 2002 (in Portuguese).
- [19] Torija MJ, Mateo E, Guillamón JM, Mas A. Identification and quantification of acetic acid bacteria in wine and vinegar by TaqMan–MGB probes. *Food Microbiol*. 2010;27(2):257–65. 10.1016/j.fm.2009.10.001
- [20] Trček J, Barja F. Updates on quick identification of acetic acid bacteria with a focus on the 16S–23S rRNA gene internal transcribed spacer and the analysis of cell proteins by MALDI-TOF mass spectrometry.
- [21] Visser't Hooft F. Biochemical studies on the genus *Acetobacter* [PhD Thesis]. Delft, the Netherlands: Delft University of Technology; 1925 (in Dutch).
- [22] Yamada Y, Aida K, Uemura T. Distribution of ubiquinone 10 and 9 in acetic acid bacteria and its relation to the classification of genera *Gluconobacter* and *Acetobacter*, especially of so-called intermediate strains. *Agric Biol Chem*. 1968;32(6):786–8. 10.1271/bbb1961.32.786

- [23] Yamada Y, Aida K, Uemura T. Enzymatic studies on the oxidation of sugar and sugar alcohol. V. Ubiquinone of acetic acid bacteria and its relation to classification of genera *Gluconobacter* and *Acetobacter*, especially of the so-called intermediate strains. *J Gen Appl Microbiol.* 1969;15(2):181–96. 10.2323/jgam.15.181
  - [24] Yamada Y, Kondo K. *Gluconoacetobacter*, a new subgenus comprising the acetate-oxidizing acetic acid bacteria with ubiquinone-10 in the genus *Acetobacter*. *J Gen Appl Microbiol.* 1984;30(4):297–303. 10.2323/jgam.30.297
  - [25] Yetiman AE, Kesmen Z. Identification of acetic acid bacteria in traditionally produced vinegar and mother of vinegar by using different molecular techniques. *Int J Food Microbiol.* 2015;204:9–16. 10.1016/j.ijfoodmicro.2015.03.013
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