

Phytochemical Screening And Antioxidant Activity Of Euryale Ferox Leaf Extract: An Untapped Botanical Resource

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ABSTRACT

Oxidative stress caused by reactive oxygen species (ROS) is a major contributor to the pathogenesis of several chronic diseases. Euryale ferox, traditionally recognized for its nutritional and medicinal properties, has been extensively studied for its seeds, while its leaves remain largely unexplored. This study investigates the antioxidant potential of E. ferox leaf extract using the 2,2-diphenyl-1-picrylhydrazyl (DPPH) free radical scavenging assay. Leaves were collected, shade-dried, and extracted with 70% ethanol. Antioxidant activity was evaluated by measuring DPPH radical scavenging, with ascorbic acid as the reference standard. The extract exhibited a concentration-dependent free radical scavenging effect. The IC50 value of the E. ferox leaf extract was found to be 71.18 \pm 0.0691 μ g/mL compared to 19.21 \pm 0.051 μ g/mL, for ascorbic acid, indicating moderate to strong antioxidant activity. These results suggest that E. ferox leaves contain significant levels of polyphenolic and flavonoid compounds responsible for their antioxidant activity, supporting their potential use as a natural antioxidant in pharmaceutical and nutraceutical formulations.

Keywords: Euryale ferox, DPPH, antioxidant, leaf extract, polyphenols, radical scavenging

1. INTRODUCTION

Oxidative stress results from an overabundance of reactive oxygen species (ROS) and inadequate antioxidant defence and significantly contributes to the development of various chronic and degenerative disorders (1). These encompass cardiovascular diseases, diabetes mellitus, cancer, rheumatoid arthritis, and neurodegenerative disorders such as Alzheimer's and Parkinson's diseases (2,3). Reactive oxygen species (ROS), such as superoxide anions, hydroxyl radicals, and hydrogen peroxide, are generated during standard cellular metabolic activities. However, when their accumulation exceeds physiological levels, it leads to lipid peroxidation, DNA damage, protein oxidation, and ultimately, cell death. Therefore, the identification and utilization of compounds that can effectively scavenge free radicals are of significant interest in preventive medicine and therapeutic intervention (4,5).

In recent years, plant-based antioxidants have garnered widespread attention due to their effectiveness, minimal side effects, and environmentally friendly nature. Phytochemicals such as flavonoids, polyphenols, alkaloids, and tannins have demonstrated significant antioxidant activity by virtue of their hydrogen-donating and electron-transfer capabilities (6,7). As part of ongoing efforts to explore novel sources of natural antioxidants, traditionally used medicinal and edible plants are being extensively investigated for their phytochemical and pharmacological potential (8,9).

Euryale ferox, known as fox nut, gorgon nut, or makhana, is an aquatic plant within the Nymphaeaceae family. It is widely cultivated in the wetlands of South and Southeast Asia, particularly in the Indian subcontinent and China (10,11). The seeds of E. ferox have been consumed for centuries as a nutritious food and are an integral component of Ayurvedic and Traditional Chinese Medicine systems. Traditionally, they have been used to manage kidney disorders, reproductive issues, and inflammatory conditions. Nutritional analysis reveals that E. ferox seeds are rich in protein, starch, dietary fiber, essential amino acids, and micronutrients such as magnesium and potassium (4). More importantly, recent studies have identified a range of bioactive compounds in E. ferox, including gallic acid, catechin, chlorogenic acid, and ferulic acid, which are known for their antioxidant properties (12,13).

The 2,2-diphenyl-1-picrylhydrazyl (DPPH) Radical scavenging test is among the most reliable and extensively utilized in vitro methods for evaluating antioxidant efficacy. This method assesses a compound's or extract's ability to contribute

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hydrogen atoms or electrons, which neutralizes DPPH free radicals (14,15). This process leads to a measurable decrease in absorbance, allowing for quantitative analysis. The simplicity, speed, and reproducibility of this assay make it a valuable tool for the preliminary screening of antioxidant capacity in plant extracts (16).

Given the traditional use of Euryale ferox and its phytochemical richness, it is imperative to scientifically validate its antioxidant potential using standardized in vitro assays. This study is meant to assess the scavenging of free radicals activity of E. ferox seed extract through the DPPH assay, to substantiate its use as a natural antioxidant. The findings may support the development of E. ferox-based formulations for use in functional foods, nutraceuticals, and complementary therapeutic systems (17,18)

2. Materials and Methods

2.1 Plant Material Collection

Fresh Euryale ferox leaves were collected from Madhubani, Biha

2.2 Preparation of Extract

The leaves were shade-dried and then ground into a coarse powder. This powdered material was extracted using 70% ethanol in a Soxhlet apparatus for 8 hours. After extraction, the mixture was filtered and concentrated. The dried extract was preserved at 4°C until it was analyzed it was analyzed (19,20).

2.3. Preliminary Phytochemical Investigation

Phytochemicals are naturally occurring bioactive compounds that contribute to the therapeutic effects of medicinal plants. The current study aimed to conduct a **preliminary phytochemical screening** of *Euryale ferox* Salisb. leaf extract prepared using 70% ethanol to identify major classes of secondary metabolites that may contribute to its biological activity (21).

Alkaloids: Detected using Mayer's and Dragendorff's reagents.

Flavonoids: Identified via the alkaline reagent and Shinoda tests. **Tannins and Phenols**: Tested using 5% ferric chloride solution.

Saponins: Evaluated using the froth formation test. **Glycosides**: Determined by the Keller–Kiliani test.

Steroids and Terpenoids: Identified using Liebermann-Burchard and

Salkowski tests.

Carbohydrates: Confirmed by Molisch's test.

Proteins: Assessed using Biuret's test.

Observations were recorded based on colour changes, precipitate formation, and frothing behaviour.

2.4 Chemicals and Reagents

All chemicals used in this study were of analytical grade. DPPH (2,2-diphenyl-1-picrylhydrazyl) was obtained from SRL Chemicals (Cat. No. SR-29128), and methanol was procured from SD Fine-Chem Limited (Cat. No. 10930lC250). Ascorbic acid, used as the reference antioxidant, was also purchased from SRL (Cat. No. 23006).

2.5 Sample Preparation

The leaves of *Euryale ferox* were collected, shade-dried, and pulverized into a fine powder. The powdered material was subjected to extraction using 70% ethanol. The resulting extract was concentrated and stored at 4°C until further use. Different concentrations of the extract were prepared in ethanol for the assay.

2.6 DPPH Radical Scavenging Assay

The antioxidant capacity of the leaf extract was evaluated using the DPPH free radical scavenging method. The assay was conducted in a 96-well microplate. To each well, $10~\mu L$ of the test extract or standard (ascorbic acid) was added to $200~\mu L$ of freshly prepared 0.1 mM DPPH solution in methanol. All samples and standards were analyzed in quadruplicate to ensure reliability. Additionally, blank wells were prepared using $200~\mu L$ of methanol and $10~\mu L$ of the test solution, but without DPPH (22,23).

Wells containing only DPPH and methanol were considered the negative control. Another control set was included, where $20~\mu L$ of deionized water replaced the test solution. The plate was kept in the dark at room temperature for 30 minutes to allow the reaction to proceed without light interference. After incubation, absorbance was measured at 517 nm using a microplate reader (iMark, Bio-Rad) (24,25).

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2.7 Data Analysis

The free radical scavenging activity was expressed as the percentage of inhibition compared to the control using the following formula:

Calculations

% RSA = ((Abs Control- Abs Sample)/Abs Control) ×100

RSA = Radical Scavenging Activity

Abs Control = Absorbance of control

Abs Sample = Absorbance of sample

The IC₅₀ value, which represents the concentration required to inhibit 50% of the DPPH radicals, was calculated using GraphPad Prism version 5. A graph was plotted with concentration on the X-axis and percent inhibition on the Y-axis to estimate the IC₅₀ values.

3. Results

3.1. Preliminary Phytochemical Screening

The phytochemical analysis of *Euryale ferox* leaf extract revealed the presence of various bioactive constituents. The results of standard qualitative chemical tests are summarized in Table 1.

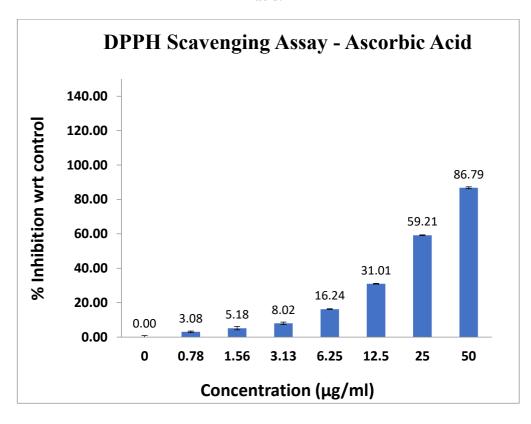
Table 1: Preliminary Phytochemical Screening of Euryale ferox Leaf Extract

3.2. DPPH Radical Scavenging Activity

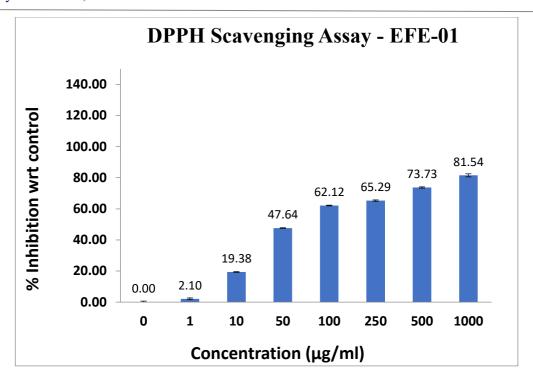
The Euryale ferox leaf extract demonstrated a concentration-dependent scavenging of DPPH radicals. As the extract concentration increased from $10 \mu g/mL$ to $100 \mu g/mL$, the percentage inhibition of DPPH increased significantly.

The IC₅₀ (half-maximal inhibitory concentration) of the *E. ferox* leaf extract was calculated to be $71.18 \pm 0.0691~\mu g/mL$ compared to $19.21\pm0.051~\mu g/mL$ for ascorbic acid.

Figure 1 Graph showing % inhibition of DPPH radicals vs. concentration of *Euryale ferox* extract and ascorbic acid.



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4. Discussion

The findings of this study provide evidence of the antioxidant potential of *Euryale ferox* leaf extract, as demonstrated by its free radical scavenging activity in the DPPH assay. The extract showed a strong dose-dependent inhibition of DPPH radicals, with an IC₅₀ value of 71.18 \pm 0.0691 μ g/mL, indicating moderate to strong antioxidant activity when compared with the standard antioxidant, ascorbic acid (18).

The initial phytochemical analysis indicated the presence of flavonoids, phenolics, alkaloids, and glycosides in the extract. These types of chemicals are recognized for their capacity to transfer hydrogen or electrons which is the primary mechanism in neutralizing free radicals like DPPH. Phenolic compounds, in particular, are effective in stabilizing free radicals due to the resonance-stabilized structure of their aromatic rings. The presence of saponins and terpenoids may also enhance the antioxidant effect through synergistic action with polyphenols (26).

The comparatively higher IC₅₀ value of the leaf extract than that of ascorbic acid may be due to the presence of a mixture of antioxidant compounds, which individually may be less potent but collectively contribute to the extract's efficacy. Furthermore, crude extracts often show reduced potency due to the presence of non-active constituents or antagonistic interactions (27).

These results suggest that *Euryale ferox* leaves can serve as a natural and accessible source of antioxidants, which could be beneficial in managing oxidative stress-related conditions, especially in neonatal health where oxidative injury can contribute to complications such as bronchopulmonary dysplasia, retinopathy of prematurity, and periventricular leukomalacia

REFERENCES

- 1. Sharifi-Rad M, Anil Kumar N V., Zucca P, Varoni EM, Dini L, Panzarini E, et al. Lifestyle, Oxidative Stress, and Antioxidants: Back and Forth in the Pathophysiology of Chronic Diseases. Front Physiol. 2020;11(July):1–21.
- 2. Pooja G, Shweta S, Patel P. Oxidative stress and free radicals in disease pathogenesis: a review. Discov Med [Internet]. 2025;2(1). Available from: https://doi.org/10.1007/s44337-025-00303-y
- 3. Holmstrup P, Damgaard C, Olsen I, Klinge B, Flyvbjerg A, Nielsen CH, et al. Comorbidity of periodontal disease: two sides of the same coin? An introduction for the clinician. J Oral Microbiol. 2017;9(1):1332710.
- 4. Jena AB, Samal RR, Bhol NK, Duttaroy AK. Cellular Red-Ox system in health and disease: The latest update. Biomed Pharmacother [Internet]. 2023;162:114606. Available from: https://www.sciencedirect.com/science/article/pii/S0753332223003943
- 5. Jomova K, Raptova R, Alomar SY, Alwasel SH, Nepovimova E, Kuca K, et al. Reactive oxygen species, toxicity, oxidative stress, and antioxidants: chronic diseases and aging. Arch Toxicol. 2023 Oct;97(10):2499–574.

- 6. Ullah A, Munir S, Badshah SL, Khan N, Ghani L, Poulson BG, et al. Important Flavonoids and Their Role as a Therapeutic Agent. Molecules [Internet]. 2020;25(22). Available from: https://www.mdpi.com/1420-3049/25/22/5243
- 7. . Maury GL, Rodríguez DM, Hendrix S, Arranz JCE, Boix YF, Pacheco AO, et al. Antioxidants in Plants: A Valorization Potential Emphasizing the Need for the Conservation of Plant Biodiversity in Cuba. Antioxidants (Basel, Switzerland). 2020 Oct;9(11).
- 8. Farag RS, Abdel-Latif MS, Abd El Baky HH, Tawfeek LS. Phytochemical screening and antioxidant activity of some medicinal plants' crude juices. Biotechnol Reports [Internet]. 2020;28:e00536. Available from: https://www.sciencedirect.com/science/article/pii/S2215017X20306202
- 9. Xu D-P, Li Y, Meng X, Zhou T, Zhou Y, Zheng J, et al. Natural Antioxidants in Foods and Medicinal Plants: Extraction, Assessment and Resources. Int J Mol Sci. 2017 Jan;18(1).
- 10. Kumar A, Yadav PK, Paras N, Nath S, Kumari U. Euryale ferox Salisb. is an Immense Potential Aquatic Cash Fruit Crop: A Review. Int J Adv Agric Sci Technol. 2016;3(3):35–49.
- 11.. Das S, Der P, Raychaudhuri U, Maulik N, Das DK. The effect of Euryale ferox (Makhana), an herb of aquatic origin, on myocardial ischemic reperfusion injury. Mol Cell Biochem. 2006 Sep;289(1–2):55–63.
- 12.. Gogoi P, Sharma P, Goudar G, Mahajan A, Ananthan R, Singh M, et al. Evaluating free-bound polyphenols, flavonoids and antioxidant properties of pigmented maize landraces. Food Chem Adv [Internet]. 2025;7:101006. Available from: https://www.sciencedirect.com/science/article/pii/S2772753X25001224
- 13. AL Zahrani NA, El-Shishtawy RM, Asiri AM. Recent developments of gallic acid derivatives and their hybrids in medicinal chemistry: A review. Eur J Med Chem [Internet]. 2020;204:112609. Available from: https://www.sciencedirect.com/science/article/pii/S022352342030581X
- 14.. Shahidi F, Zhong Y. Measurement of antioxidant activity. J Funct Foods [Internet]. 2015;18:757–81. Available from: https://www.sciencedirect.com/science/article/pii/S1756464615000511
- 15. Ismail R, Tannous JH. Investigation of the free radical stability of 2,2-diphenyl-1-picrylhydrazyl upon storage in hydrogen- and electron–donning solvents at different temperatures. Case Stud Chem Environ Eng [Internet]. 2025;11:101207. Available from: https://www.sciencedirect.com/science/article/pii/S2666016425001148
- 16.. Patil ND, Bains A, Sridhar K, Sharma M, Dhull SB, Goksen G, et al. Recent advances in the analytical methods for quantitative determination of antioxidants in food matrices. Food Chem [Internet]. 2025;463:141348. Available from: https://www.sciencedirect.com/science/article/pii/S0308814624029984
- 17.. Wu C, Chen R, Wang XS, Shen B, Yue W, Wu Q. Antioxidant and anti-fatigue activities of phenolic extract from the seed coat of Euryale ferox Salisb. and identification of three phenolic compounds by LC-ESI-MS/MS. Molecules. 2013 Sep;18(9):11003–21.
- 18. Lee SE, Ju EM, Kim JH. Antioxidant activity of extracts from Euryale ferox seed. Exp Mol Med. 2002 May;34(2):100–6.
- 19.. Dhore SB, Ghatpande KS, Barabole AH, Ahire RS. EXPLORING THE DIVERSE ACTIVITIES OF VITEX NEGUNDO: A COMPREHENSIVE. 2024;12(4):49–54.
- 20. Kiranmai VS, Kumar PK, Rajesham V V., Rao TR. Extraction Methods and Pharmacological Importance of Clitoria ternatea Plant. Int J Pharm Sci Rev Res. 2023;83(2):99–105.
- 21.. Devi OR, Moirangthem L, Keithellakpam OS, Sharma N, Singh KB. Phytochemical Screening and Evaluation of Antioxidant Potential in Euryale ferox Salisb. and Eupatorium birmanicum DC. of Manipur, India. Pharmacogn J. 2024 Dec;16:1231–7.
- 22. Baliyan S, Mukherjee R, Priyadarshini A, Vibhuti A, Gupta A, Pandey RP, et al. Determination of Antioxidants by DPPH Radical Scavenging Activity and Quantitative Phytochemical Analysis of Ficus religiosa. Molecules. 2022 Feb;27(4).
- 23.. Bibi Sadeer N, Montesano D, Albrizio S, Zengin G, Mahomoodally MF. The Versatility of Antioxidant Assays in Food Science and Safety-Chemistry, Applications, Strengths, and Limitations. Antioxidants (Basel, Switzerland). 2020 Aug;9(8).
- 24.. Guleria S, Tiku AK, Koul A, Gupta S, Singh G, Razdan VK. Antioxidant and antimicrobial properties of the essential oil and extracts of Zanthoxylum alatum grown in north-western Himalaya. ScientificWorldJournal. 2013;2013:790580.
- 25. Di Bella CE, Censi R, Zara S, Di Martino P, Gigliobianco MR. Activation of polyphenolic compounds as new ingredients for sustainable cosmetic formulation. J Drug Deliv Sci Technol [Internet]. 2025;108:106918. Available from: https://www.sciencedirect.com/science/article/pii/S1773224725003211

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- 26.. Phytochemical Screening and Quantitative Determination of Phytochemicals in Leaf Extracts of Hannoa undulata. Int J Med Plants Nat Prod. 2018;4(2):32–8.
- 27. Singh R, Sharma S, Sharma V. Comparative and quantitative analysis of antioxidant and scavenging potential of Indigofera tinctoria Linn. extracts. J Integr Med. 2015 Jul;13(4):269–78..

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